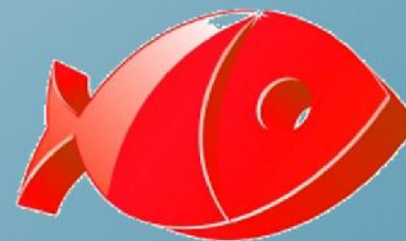
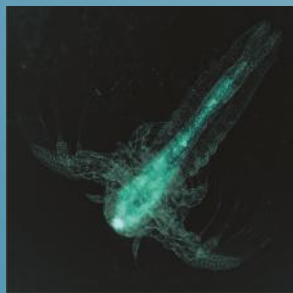
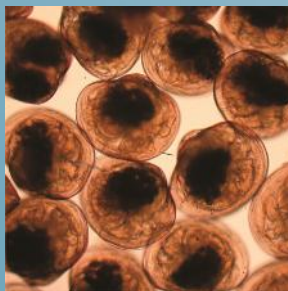
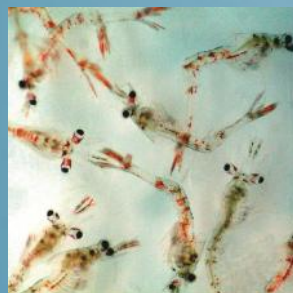
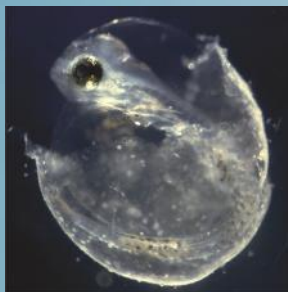


# larvi 2013

6th fish & shellfish larviculture symposium

Bacterial community assembly in  
developing cod larvae (*Gadus morhua*)

Ingrid Bakke



ghent university, belgium, 2-5 september 2013

# Bacterial community assembly in developing cod larvae (*Gadus morhua*)



Photo: Akvaplan-Niva AS

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# Introduction



Photo: LarvalBase

- After hatching and mouth opening, the intestinal system of a fish larva is colonized
- This colonization is important to host health and development
- Conserved responses to colonization of the gut system (mouse and zebra fish):
  - Stimulation of epithelial cell proliferation
  - Promotion of nutrient metabolism
  - Development of the mucosal immune system
- Microbiota associated with cultivated marine fish larvae
  - The larvae shear the rearing water with bacteria
  - The digestive and immune systems are immature
  - Vulnerable to opportunistic bacteria
- Possible to steer the larval microbiota?

# Background

- Need for knowledge:
  - Sources for the larval microbiota?
  - **Determinants for the composition of the gastrointestinal microbiota?**

Results for cod larvae in PROMICROBE at NTNU/SINTEF in Trondheim:

- Study I: Effect of different live feed diets on larval microbiota
- Study II: Effect of different rearing water systems on larval microbiota
- Study III: A deep sequencing approach to characterize variation in larval microbiota between individuals and with time



Photo: Nofima

# Study I Effect of live feed diet

## Experimental design

- First feeding experiment with cod larvae
- Three different live feed diets from 3 to 22 dph:
  - Diet **COP**: Copepods cultured on microalgae *R. baltica*
  - Diet **RR**: Rotifers cultured on microalgae *R. baltica*
  - Diet **CR**: Rotifers cultured on Bakers yeast and MarolE
- For each diet: Three tanks, totally 9 tanks (100 l)
- From 18 – 36 dph: Artemia to all tanks
- Microbial communities investigated for individual larvae, water and live feed samples by PCR/DGGE



Experimental tanks at SINTEF Fisheries and Aquaculture

environmental  
microbiology reports

Environmental Microbiology Reports (2013) 5(4), 537–548



doi:10.1111/1758-2229.12042

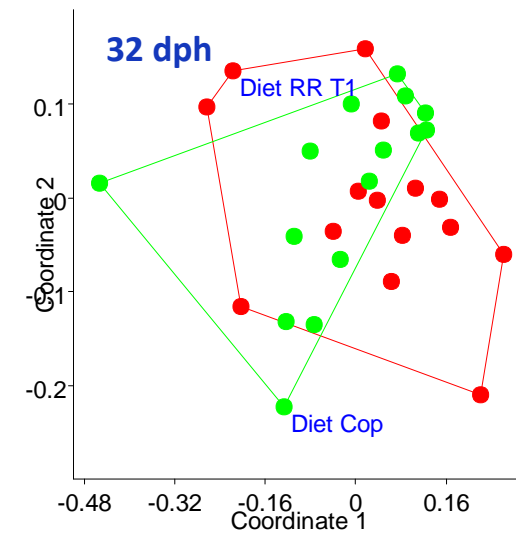
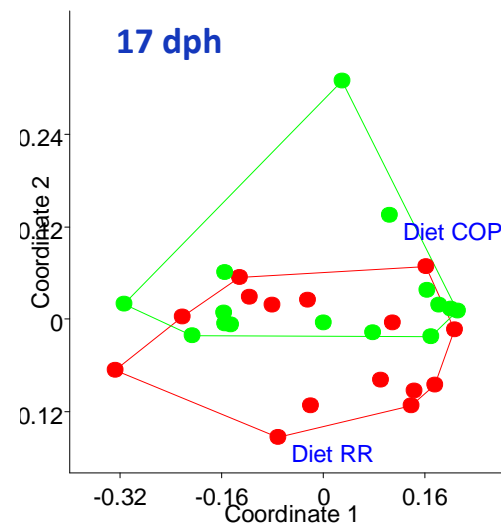
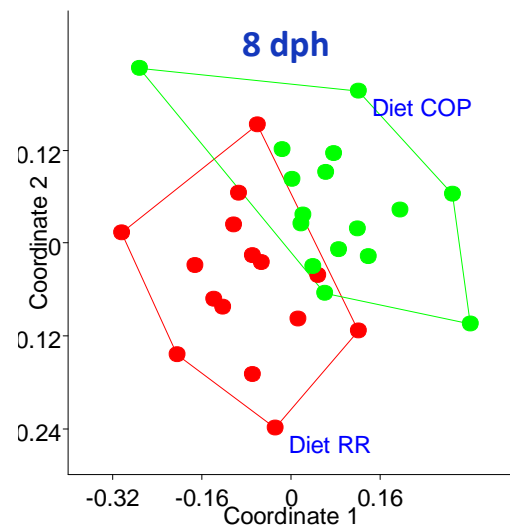
**Live feed is not a major determinant of the microbiota associated with cod larvae (*Gadus morhua*)**

Ingrid Bakke,<sup>1\*</sup> Jorunn Skjermo,<sup>3</sup> Tu Anh Vo<sup>2</sup> and Olav Vadstein<sup>1</sup>

# Study I Results

- The live feed diets represented different microbiotas
- Small, but significant differences in larval microbiota between all diets at 8 dph
- No significant differences in larval microbiota between diets at 17 and 32 dph

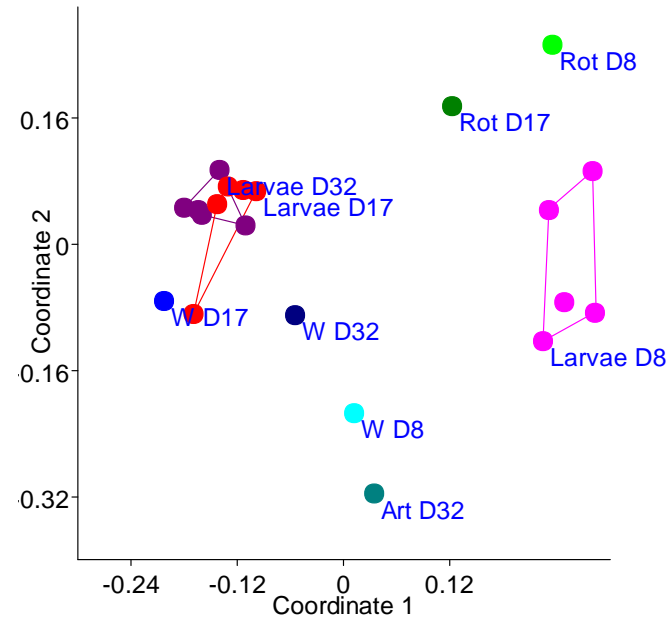
Non-metric MDS based on Bray-Curtis similarities for cod larval microbiota reared with RR and COP diets



# Study I Results

- At 17 and 32 dph:  
No differences in larval microbiota due to rearing with different diets  
Differences in larval microbiota due to rearing in different tanks
- Larval microbiota at 17 and 32 dph remarkable similar despite change of diet
- Larval microbiota generally more similar to water microbiota than to live feed microbiota

RR Tank 3: Larval, water and live feed microbiota



# Study II Effect of rearing water

## Experimental design

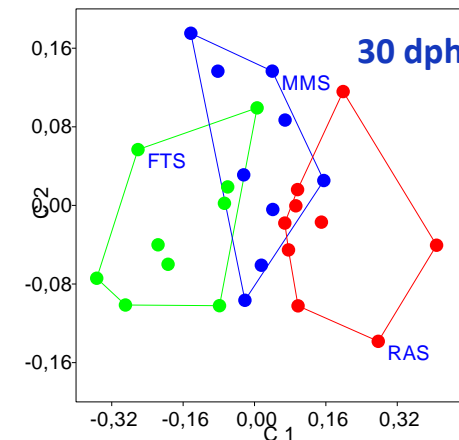
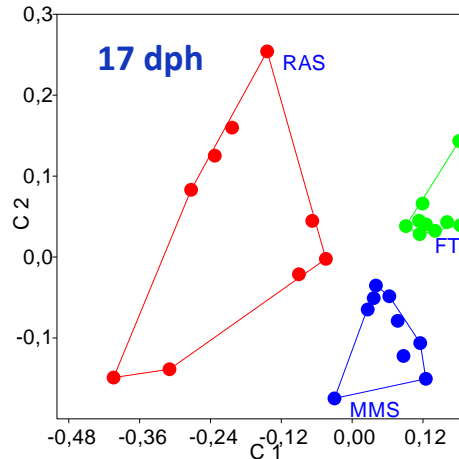
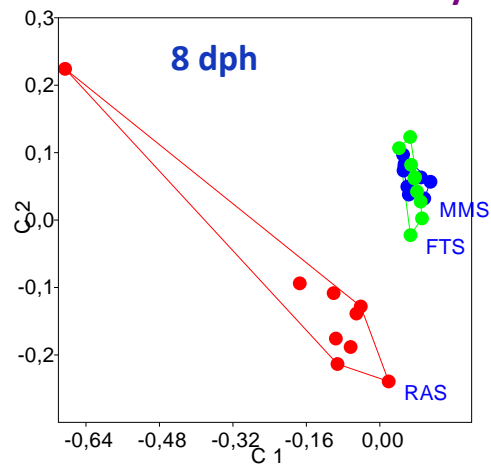
- First feeding experiment with cod larvae
- Three different rearing water systems until 30 dph:
  - FTS:** Conventional flow-through system
  - MMS:** Microbially matured water (with biofilter) in a flow-through system
  - RAS:** Recirculating aquaculture system without disinfection
- For rearing water system: Three tanks, totally 9 tanks (100 l)
- After 30 dph: All tanks received MMS water
- Microbial communities investigated for individual larvae, water and live feed samples by PCR/DGGE



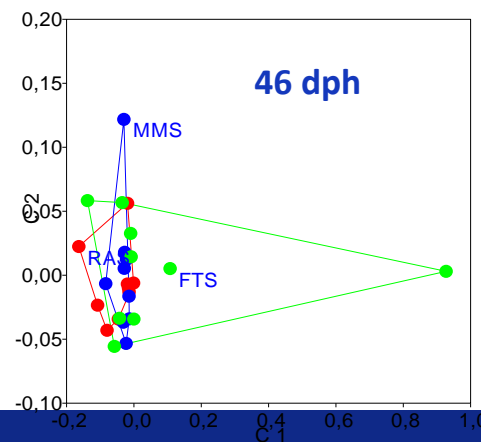
# Study II Results

- Significant differences in larval microbiota between different water rearing systems
- Change of identical water (MMS) to all tanks → similar larval microbiota between tanks

Non-metric MDS based on Bray-Curtis similarities for cod larval microbiota reared with FTS, MMS, and RAS

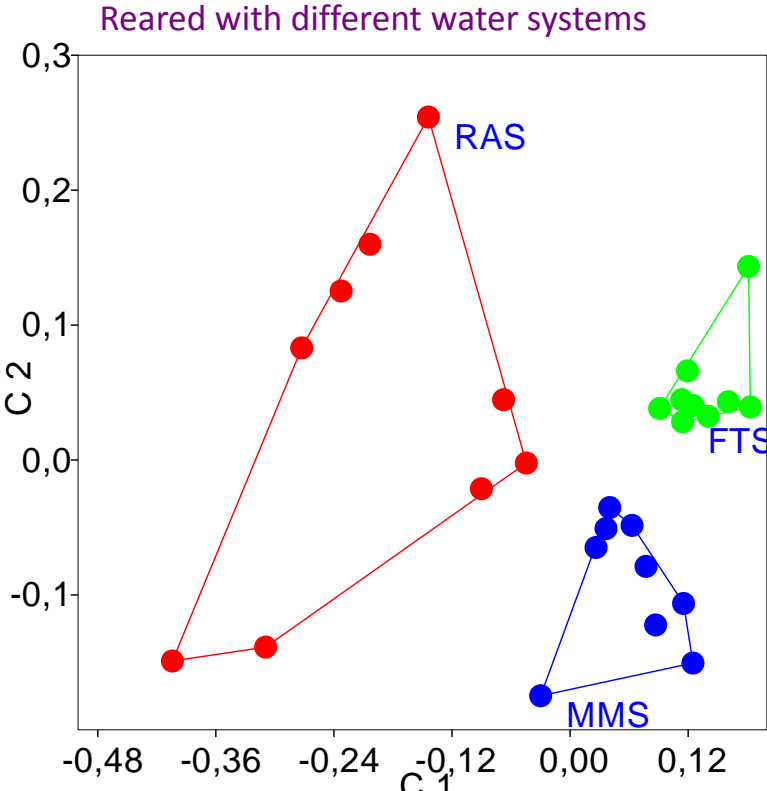
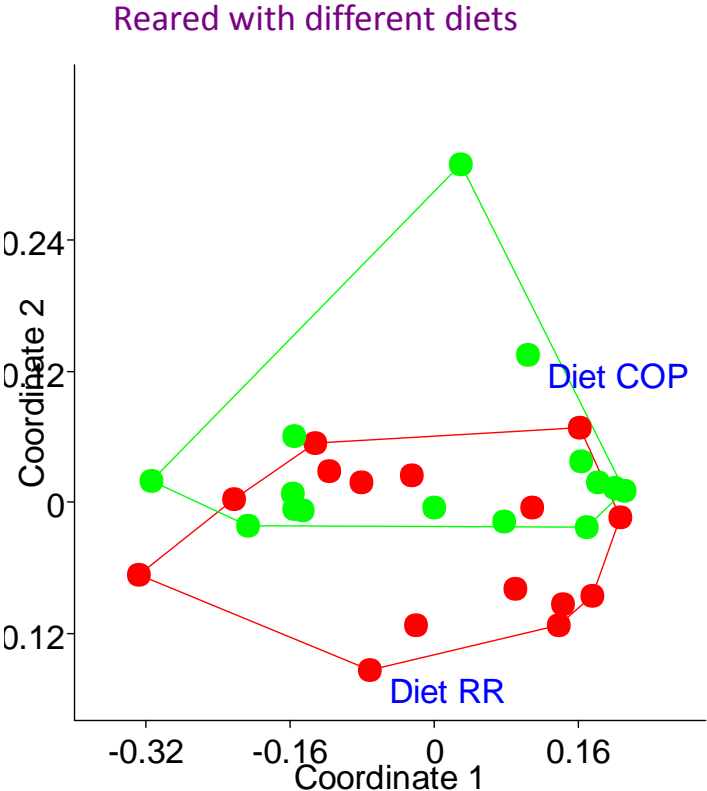


All tanks received MMS water



# Study I versus Study II

## Larval microbiota 17 dph:



# Study III

## Deep seq. of microbiota associated with developing cod larvae

### Experimental design

- Microbial communities characterized by amplicon pyrosequencing (“deep sequencing”)
- Two tanks from Study I (the “diet study”): diets from 3 to 22 dph:
  - Tank 1 (T1) Diet **COP**: Copepods cultured on microalgae *R. baltica*
  - Tank 2 (T2) Diet **RR**: Rotifers cultured on microalgae *R. baltica*
- Target sequence: Variable region 4 of the bacterial 16S rRNA gene

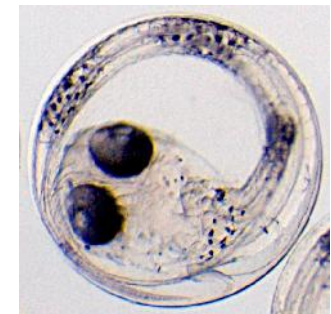
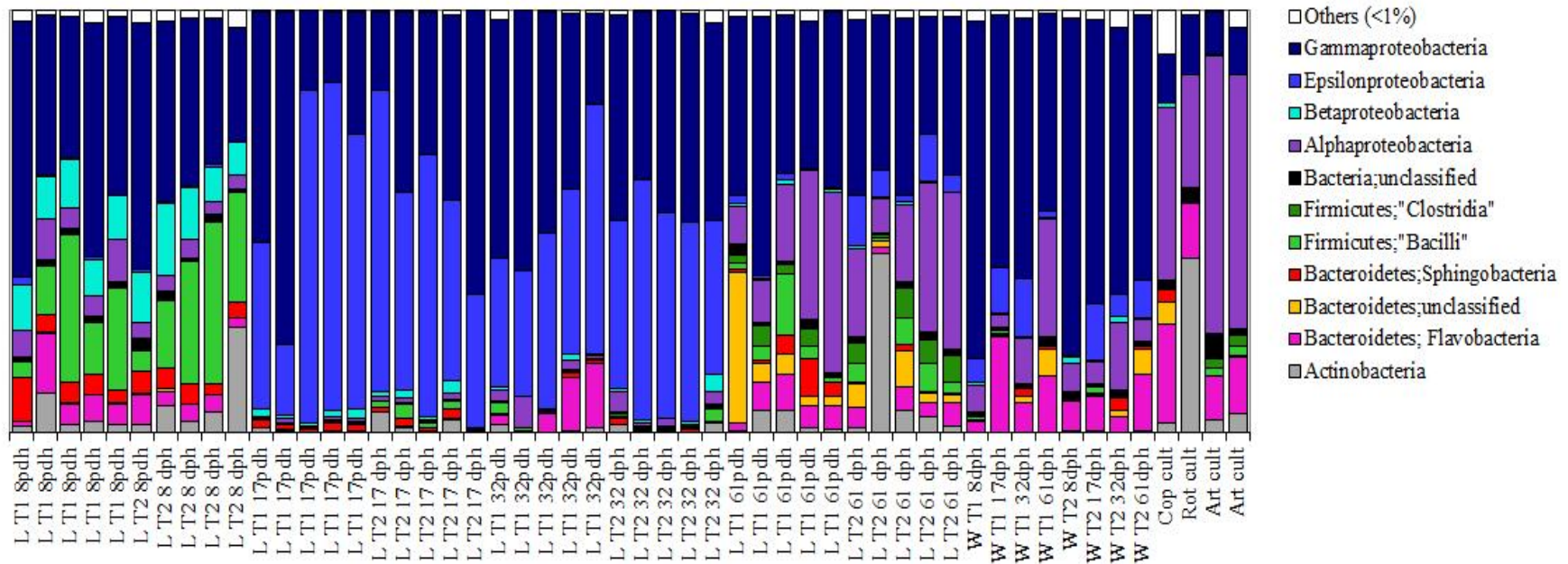


Photo: Tora Bardal

# Study III Results

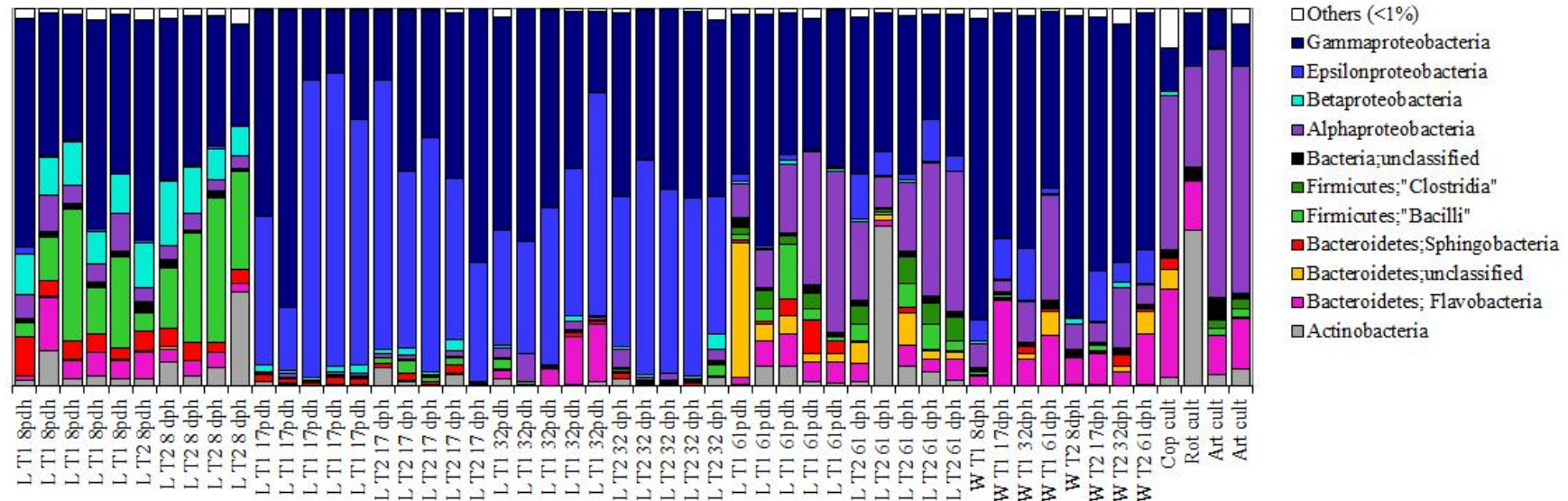
- Large number of high quality DNA sequences were obtained  
Average number of sequences (“after trimming”) per sample:  
Larva: 6248 ± 1196  
Rearing water: 8424 ± 514  
Live feed: 8580 ± 649



# Study III Results

## Larval microbiota: Temporal trends

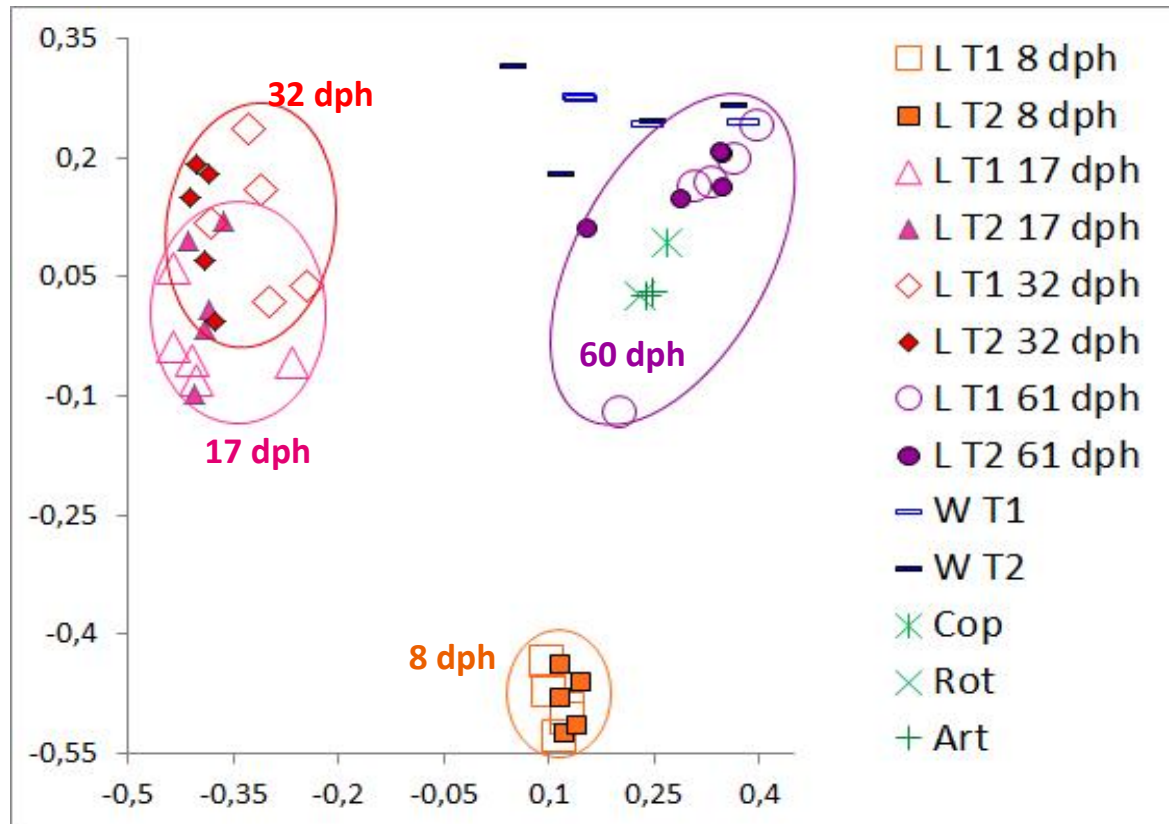
- The composition of the larval microbiota changes with age:
  - 8 dph: Pseudomonas, diverse  $\beta$ -proteobacteria, Bacilli is abundant
  - 17 dph and 32 dph: Low diversity, arcobacter ( $\epsilon$ -proteobacteria) and  $\gamma$ -proteobacteria
  - 61 dph: High diversity, diverse  $\gamma$ -proteobacteria, high abundance of Rhodobacter



# Study III Results

- Similar larval microbiota at 17 and 32 dph
- No differences in larval microbiota between Tank 1 and Tank 2 (COP and RR diets)

Pco plot based on Bray-Curtis distances for T1 (COP) and T2 (T2)

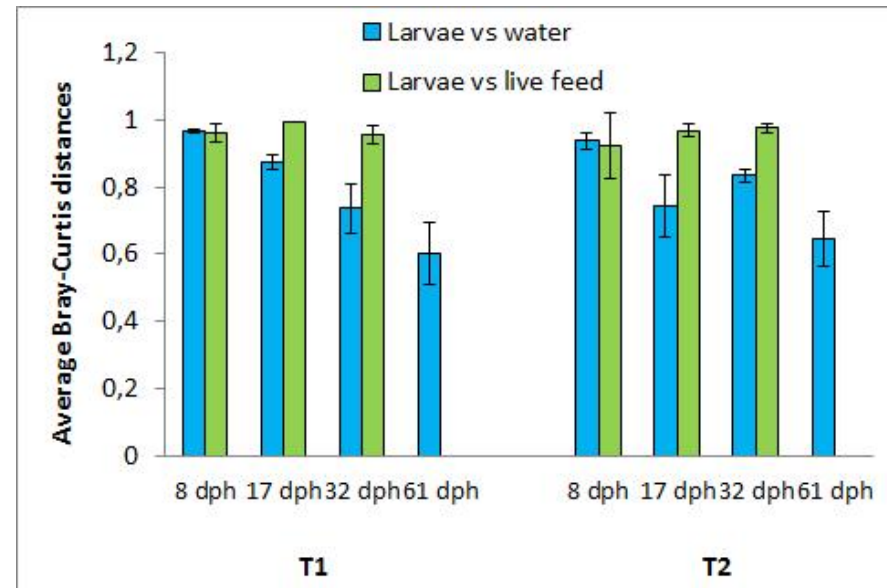


# Study III Results

Comparison of larval microbiota to water and live feed microbiota

- Larval microbiota highly dissimilar from live feed microbiota
- Larval microbiota highly dissimilar from water microbiota

Higher similarity with increasing age (excretion?)



- Where do the bacteria associated with the 8 dph larvae come from?
  - The most abundant larval OTUs are rare in the water and live feed microbiotas

OTU classification	% abundance Tank 1			% abundance Tank 2		
	Larvae	Water	COP Feed	Larvae	Water	Rot Feed
Pseudomonas	29.8	0.05	0.09	26.3	0.05	0.06
Bacillales	13.1	0.01	-	16.5	-	0.01
Microbacterium	3.6	0.18	0.30	6.4	0.38	40.5
Enterobacteriaceae	4.2	-	0.01	3.9	-	-

# Summary

- Live feed diet seems to have little influence on the larval microbiota
- Rearing water affects the larval microbiota: Rearing water the major source for larval microbiota?
- Major changes in community structure during larval development
- The early larval microbiota very different from water and live feed microbiota: strong host selection?

# Questions

- Determinants for community structure of the larval microbiota, what cause the changes in community structure?
  - Developmental changes in the digestive system?
  - Developmental changes in the immune system?
- Differences in early larval microbiota when rearing with different water systems:  
Which bacteria make these differences?  
Pyrosequencing of samples from Study II may provide an answer



# Thank you for your attention!



This work was funded by The European Community's Seventh Framework Programme (FP7/2007–2013) under grant agreement no. 227197 Promicrobe 'Microbes as positive actors for more sustainable aquaculture'.